

R.G.C.C.- RESEARCH GENETIC CANCER CENTRE S.A.

Florina, 20.02.2024

Dear Colleague,

We report the allelic discrimination results for patient John Doe whose sample receipt on 08.02.2024. DNA was extracted from blood sample and was used as template in PCR reactions. Molecular-based assays and spectrophometer analysis were used to verify the DNA. In all reactions genomic DNA was used as a positive control. The reactions were performed in triplicates.

The graduated bars indicate any potential positive or negative outcome. An arrow on the green or the red part of the bar demonstrates the outcome.



Basic

| Drug | Polymorphism | Outcome |
|----------|------------------|-----------------------------|
| | CYP1A1*2C | ∇ Normal Metabolizer |
| | CYP1A2*1F | Possible Normal Metabolizer |
| | CYP1A2*1K | Normal Metabolizer |
| | CYP1B1*Leu432Val | Possible Poor Metabolizer |
| | CYP2C19*17 | Fast Metabolizer |
| | CYP2C19*2 | Normal Metabolizer |
| N. I | CYP2C19*3 | Poor Metabolizer |
| Phase I | CYP2C9*2 | Normal Metabolizer |
| | CYP2C9*3 | Normal Metabolizer |
| | CYP2D6*10 | |
| | CYP2D6*2 | Normal Metabolizer |
| | CYP2D6*3B | Normal Metabolizer |
| | CYP3A4*1B | Normal Metabolizer |
| | CYP3A4*20 | Normal Metabolizer |
| Phase II | ABCB1*Ile1145Ile | Possible Poor Metabolizer |
| | ABCG2*Gln141Lys | Normal Metabolizer |

| EI | PHX1*His139Arg | Normal Metabolizer |
|----|----------------|-----------------------------|
| EI | PHX1*Tyr113His | Normal Metabolizer |
| G | STP1*Ala114Val | Normal Metabolizer |
| G | STP1*Ile105Val | Normal Metabolizer |
| | NAT2*11A | Possible Normal Metabolizer |
| | NAT2*12A | Possible Normal Metabolizer |
| | NAT2*13 | Possible Normal Metabolizer |
| | NAT2*14 | Normal Metabolizer |
| | NAT2*5D | Possible Normal Metabolizer |
| | NAT2*6B | Possible Normal Metabolizer |
| | NAT2*7A | Normal Metabolizer |
| | TPMT*2 | Normal Metabolizer |
| | TPMT*4A | Normal Metabolizer |

ALKYLATING AGENTS

| Drug | Polymorphism | Outcome |
|--------------------|-------------------|--|
| Carboplatin | ALDH3A1*Pro329Ala | Increased likelihood of cystitis (carboplatin, cyclophosphamide, thiotepa) |
| | COMT*19955692C>T | Decreased risk of Ototoxicity in children |
| | ERCC1*Gln504Lys | Better response and decreased vomiting during the first 24 hours post-cisplatin administration (granisetron or palonosetron) |
| Cisplatin | GSTP1*Ile105Val | Increased risk of toxicity |
| | LRP2*Lys4094Glu | Decreased risk of hearing loss |
| | XPC*Gln902Lys | Increased risk for toxicity |
| Cyclophosphamide | ALDH3A1*Pro329Ala | Increased likelihood of cystitis (carboplatin, cyclophosphamide, thiotepa) |
| Platinum compounds | ERCC1*Asn118Asn | Increased risk for nephrotoxicity |
| | ERCC1*Gln504Lys | Increased risk for nephrotoxicity |
| | NQO1*Pro149Ser | Increased overall, progression-free survival |

Topo I Inhibitors

| Drug | Polymorphism | Outcome |
|------------|------------------|-------------------------------|
| Irinotecan | UGT1A1*172270T>G | Increased risk of neutropenia |
| | UGT1A1*Gly71Arg | Decreased risk of neutropenia |

Topo II Inhibitors

| Drug | Polymorphism | Outcome |
|----------------|------------------|---|
| Anthracyclines | CBR1*133G>A | Increased risk of cardiomyopathies (low to moderate dose) |
| | CBR3*Val244Met | Increased risk of Heart Failure |
| Daunorubicin | NRP2*110077C>G | Decreased IC50 |
| Doxorubicin | ABCC2*Cys1515Tyr | Increased risk of cardiotoxicity |
| | ABCC2*Val1188Glu | Increased risk of cardiotoxicity |

Antimetabolites

| Drug | Polymorphism | Outcome |
|----------------|----------------|---|
| | DPYD*1905+1G>A | Increased risk of drug toxicity |
| | DF1D*1903+1G>A | (5-fluorouracil, capecitabine, tegafur) |
| | DPYD*1905+1G>A | Increased risk of drug toxicity, Leukopenia, Mucositis |
| | DPYD*1905+1G>A | Increased risk of drug toxicity |
| | DPYD*1905+1G>A | Decreased metabolism |
| | DPYD*1905+1G>A | Increased likelihood of drug toxicity (5-fluorouracil, capecitabine) |
| | DPYD*1905+1G>A | Increased likelihood of Thrombocytopenia |
| | DPYD*1905+1G>A | Increased likelihood of Mucositis |
| | DPYD*Asp949Val | Decreased severity of drug toxicity |
| 5-Fluorouracil | DPYD*Asp949Val | Decreased likelihood of drug toxicity (5-fluorouracil, capecitabine) |
| | DPYD*Cys29Arg | Increased likelihood of Nausea and Vomiting |
| | DPYD*Cys29Arg | Decreased likelihood of overall gastrointestinal toxicity |
| | DPYD*Cys29Arg | Increased metabolism |
| | DPYD*Ile543Val | Decreased likelihood of Nausea and Vomiting |
| | DPYD*Ile543Val | Increased clearance |
| | DPYD*Ile543Val | Decreased likelihood of Leukopenia |
| | DPYD*Met166Val | Decreased metabolism |

| | DPYD*Met166Val | Decreased likelihood of Neutropenia |
|--------------|------------------|--|
| Capecitabine | DPYD*1905+1G>A | Increased risk of drug toxicity (5-fluorouracil, capecitabine, tegafur) |
| | DPYD*1905+1G>A | Increased likelihood of drug toxicity |
| | DPYD*Asp949Val | Decreased likelihood of drug toxicity (5-fluorouracil, capecitabine) |
| Cytarabine | CDA*20915590delC | Decreased drug toxicity |
| | CDA*-92A>G | Decreased drug toxicity |
| | CDA*Lys27Gln | Decreased drug toxicity |
| Gemcitabine | CDA*Ala70Thr | Decreased severity of Neutropenia |
| Methotrexate | MTHFR*Glu429Ala | Decreased risk of mucositis |

Spindle Poisons

| Drug | Polymorphism | Outcome |
|------------|-----------------|---------------------------------|
| Docetaxel | CYP3A4*1B | Decreased clearance |
| Paclitaxel | CYP2C8*23210C>G | Increased risk of Neurotoxicity |
| | CYP3A5*12083G>C | Decreased risk of Neurotoxicity |

Appendix:

Drug Metabolism:

Phase I:

Phase I enzymes are responsible reactions that convert parent compound into a more polar metabolite by adding or unmasking functional groups. Usually these metabolites are inactive. Phase I reactions include, oxidation, reduction, hydrolytic cleavage, alkylation, methylation, ring cyclization etc. These reactions prepare chemicals for phase II metabolisms and subsequent excretion.

The Cytochrome P450 (CYP) enzyme superfamily is the most important system in the biotransformation of many endogenous and exogenous substances, such as drugs, toxins and carcinogens. For drug metabolism the most important polymorphisms are those of the genes coding for CYP2C9, CYP2C19, CYP2D6 and CYP3A4. CYP1A1 and CYP1A2 are among the most responsible for biotransformation of chemicals, especially for the metabolic activation of precarcinogens. Genetic polymorphism is an important reason for variations in drug response of the human body. There are four distinct phenotypes: poor metaboliser (PM), intermediate metaboliser (IM), extensive metaboliser (EM) and ultrarapid metaboliser (UM). A poor metaboliser lacks active allele and may present adverse effects at usual doses, due to reduced metabolism and increased drug concentration. Individuals with intermediate metabolic phenotype are homozygous for two reduced activity alleles or are heterozygous for an inactive allele. Extensive metabolisers have two fully active allele and show the expected response to a standard dose. Ultra extensive metabolisers are individuals with more than two copies of active gene.

- Cytochrome P450 2D6 is one of the most important enzymes, involved in the metabolisms of xenobiotics in the body, but also in activation of many substances in their active compounds.
- Cytochrome P450 2C19 is responsible for metabolisation or activation of many hormones and drugs (anti-epileptics, anti-depressants, anti-platelet clopidogrel, esomeprazole).
- Cytochrome P450 1A2 is involved in metabolism of xenobiotics substrates such caffeine, aflatoxin B1 and acetaminophen.
- Cytochrome P450 3A4 is one of the most important enzymes involved in xenobiotics metabolism in human body. It metabolizes some steroids and carcinogens. Approximately half of the drugs that are used are metabolized by this protein, such acetaminophen, codeine, cyclosporine, diazepam and erythromycin.
- Cytochrome P450 2C9 is an enzyme with a major role in the oxidation of both xenobiotics and endogenous compounds. Warfarin, phenytoin, acenocoumarol, tolbutamide, losartan glipizide and a few nonsteroidal anti-inflammatory drugs (aspirin, ibuprofen, naproxen) are metabolized by CYP2C9.

Phase II:

The Phase II reactions are conjugations with endogenous substrate to further increase aqueous

solubility and conjugations with glucoronide, sulfate, acetate, amino acid etc. N-acetyltransferase 2

(NAT2), Epoxide hydrolase 1 (EPHX1), Glutathione S-transferase P (GSTP1) and Thiopurine methyltransferase (TPMT) are the major enzymes involved in phase II drug metabolism.

• N-acetyltransferase 2 (NAT2), is an enzyme that activates and deactivates arylamine and hydrazine

drugs and carcinogens. Human populations segregated into rapid, intermediate and slow acetylator

phenotypes, according to different polymorphisms combinations.

• Glutathione S-transferases are responsible for the detoxification of a range of drugs and potential

carcinogens, through glutathione conjucation. The GSTP1 is associated with xenobiotics metabolism

and susceptibility to cancer and other diseases.

• Thiopurine S-methyltransferase (TPMT) is an enzyme that metabolises thiopurine drugs such as

azathioprine, 6-mercaptopurine and 6-thioguanine. Individual homozygous for two non-functional

TPMT variants are at high risk for toxic side effects, due to decreased methylation and decreased

inactivation of 6MP.

Pharmacodynamics:

• P-glycoprotein 1, or multidrug resistance protein 1, or ATP-binding cassette sub-family B member 1

(ABCB1), or CD243, is an ATP-dependent drug efflux pump for xenobiotics compounds with broad

substrate specificity. ABCB1 regulates the distribution and bioavailability if drugs, removes toxic

metabolites and xenobiotics from cells, transports compounds out of brain and protects hematopoietic

stem cells from toxins.

• ATP-binding cassette sub-family G member 2 (ABCG2), is a xenobiotic transporter with important

role in the multidrug resistance phenotype of several cancer cell lines.

Sincerely,

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