

• Patient name: **demo123 demo1345**

• Gender: **Man**

• Date of birth: **03-02-1996**

• Sample code: **SPR00188AA**

• Sample date: **20-11-2023**

• Report date: **20-11-2023**



Fagron Sport Test

Genetic report



Fagron Sport Test

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Patient identification data

1



Patient name —●— **demo123 demo1345**
 Gender —●— **Man**
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 Sample type —●— **Buccal Swab**
 Sample code —●— **SPR00188AA**
 Sample date —●— **20-11-2023**
 Report date —●— **20-11-2023**

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Presentation of the test

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What is this test about?

This is a genetic test that analyzes DNA to evaluate 25 genetic variants with the intention of informing about predispositions and risks associated with individual athletic performance. This information should be integrated with physical (eg, age, gender, body mass index, VO2 max, etc.) and behavioral (eg, eating habits, physical activity, etc.) characteristics to establish the best personalized training plan.

Who is it addressed to?

It is a test aimed at healthcare professionals and specialists in sports medicine.

For what type of patient?

It is designed for people who practice sports, both privately and professionally. Thanks to the study of our genetics we will be able to know and evaluate our sports possibilities and also our limitations, prevent injuries, improve our performance, and ultimately take care of our health.

How do you evaluate genetic variants?

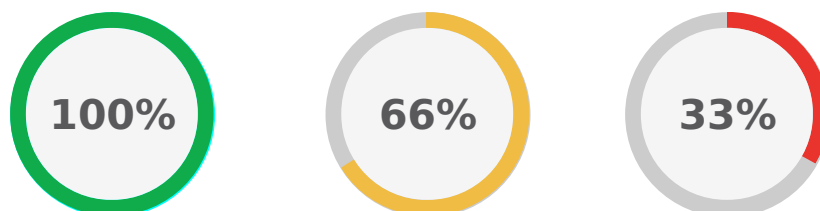
The test evaluates the genetic variants analyzed, classifying them as "**favourable**", "**intermediate**" and "**unfavourable**":



* Having an "unfavorable" genotype doesn't mean someone can't exercise normally, it just suggests that they might struggle more than someone with a more favorable genotype.

How do you evaluate the categories?

The test assigns a value to each genotype and calculates the total value per category (eg "muscular properties", "power", etc.) represented by graph.



Sports skills

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3.1. Power (anaerobic exercise)

Anaerobic exercise requires short periods of maximal muscular effort. Its practice contributes to improving muscle power, strength and size.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ACTN3	rs1815739 (C > T)	C	CT	Predisposition for power sports
AGT	rs699 (A > G)	G	AA	No predisposition for power sports
AMPD1	rs17602729 (G > A)	G	GG	Predisposition for power sports
HIF1A	rs11549465 (C > T)	T	CC	No predisposition for power sports
LRPPRC	rs10186876 (A > G)	A	AA	Predisposition for power sports
PPARG	rs1801282 (C > G)	G	CC	No predisposition for power sports
PPARGC1A	rs8192678 (C > T)	T	TT	Predisposition for power sports
SOD2	rs4880 (A > G)	A	GG	No predisposition for power sports

3.2. Endurance (aerobic exercise)

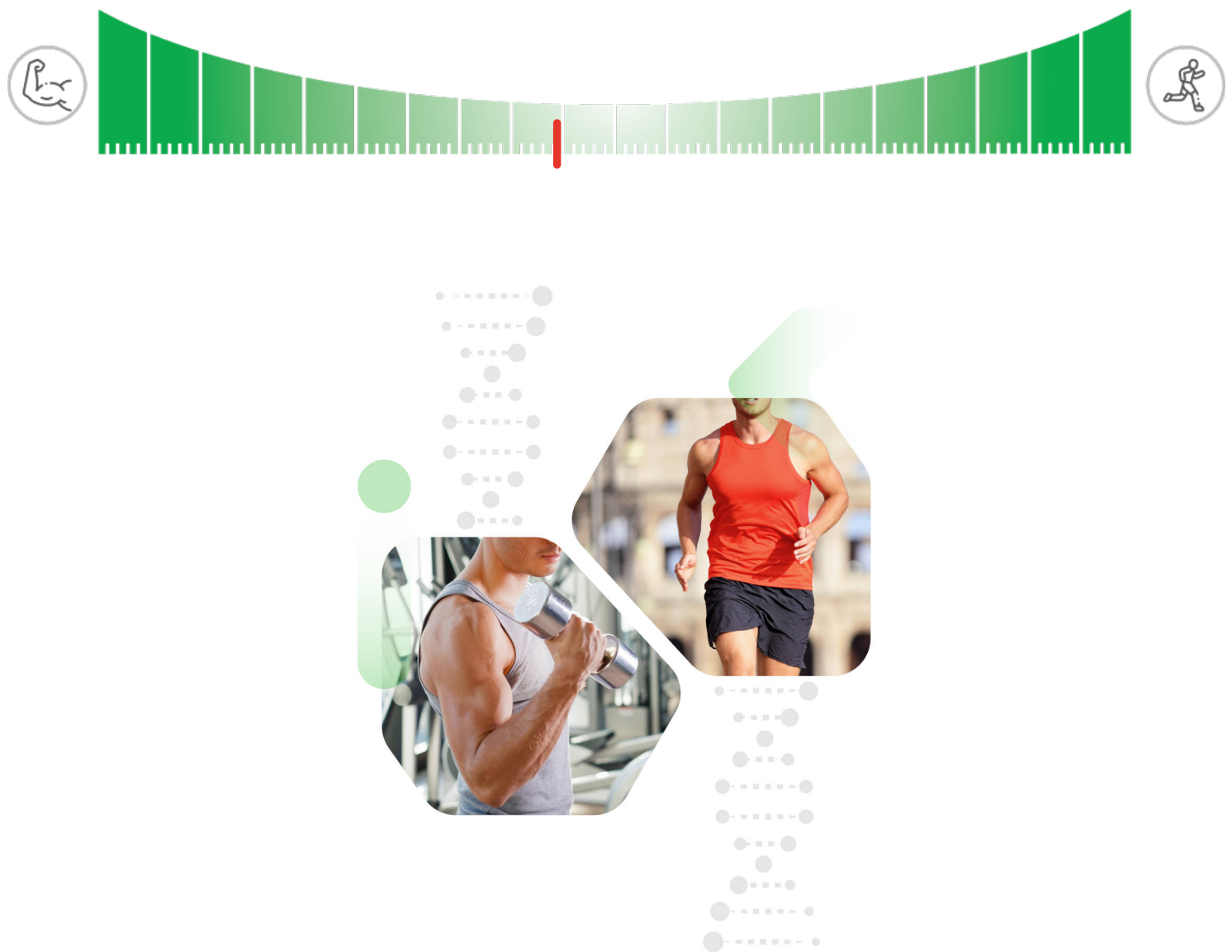
Aerobic exercise involves long periods of average muscular exertion. Its practice contributes to maintaining a healthy heart, lungs and circulatory system, improving cardiorespiratory fitness.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ACE	rs4343 (G > A)	A	GG	No predisposition for endurance sports
ADRB2	rs1042713 (G > A)	A	GA	Slight predisposition for endurance sports
AMPD1	rs17602729 (G > A)	G	GG	Predisposition for endurance sports
AQP1	rs1049305 (G > C)	C	GC	Predisposition for endurance sports
COL5A1	rs12722 (C > T)	T	TT	Predisposition for endurance sports
GSTP1	rs1695 (A > G)	G	AA	No predisposition for endurance sports
HFE	rs1800562 (G > A)	A	GG	No predisposition for endurance sports
HIF1A	rs11549465 (C > T)	C	CC	Predisposition for endurance sports
PPARG	rs1801282 (C > G)	G	CC	No predisposition for endurance sports
PPARGC1A	rs8192678 (C > T)	C	TT	No predisposition for endurance sports

3.3. Power vs. Endurance

According to the results of power and resistance, the global predisposition of the patient is:



3.4. Aerobic potential (VO2 max)

Aerobic fitness (cardiovascular endurance) is the body's ability to supply oxygen to the muscles, allowing them to work or participate in physical activity. The VO2 max test is the most accurate test of aerobic or cardiovascular fitness. Studies based on the measurement of VO2 max have identified SNPs in genes related to the development of the cardiovascular system associated with maximal gains in VO2 max from exercise.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ADRB2	rs1042713 (G > A)	A	GA	Predisposition to limited improvement in VO2 max in response to endurance training.
AMPD1	rs17602729 (G > A)	G	GG	Predisposition to improve VO2 max in response to endurance training.
GSTP1	rs1695 (A > G)	G	AA	Predisposition to very limited improvement in VO2 max in response to endurance training.
HFE	rs1800562 (G > A)	A	GG	Predisposition to very limited improvement in VO2 max in response to endurance training.
HIF1A	rs11549465 (C > T)	C	CC	Predisposition to improve VO2 max in response to endurance training.
PPARD	rs2267668 (G > A)	A	AA	Predisposition to improve VO2 max in response to endurance training.
PPARGC1A	rs8192678 (C > T)	T	TT	Predisposition to improve VO2 max in response to endurance training.
ZIC4	rs11715829 (T > C)	C	TT	Predisposition to very limited improvement in VO2 max in response to endurance training.

Calculation of the VO2 max

The best and most precise method to determine the VO2max value is a breath analysis. An alternative less precise is the Copper test, a 12-minute run that measures the maximum distance covered during that time. The VO2max value can then be determined using the following formula: **VO2max = (distance traveled in meters - 505) / 45**. You can use this table to estimate your VO2 max value using the Cooper test:

Age	Very bad	Bad	Regular	Good	Excellent	Higher
13-19	< 35,0	35,0 - 38,3	38,4 - 45,1	45,2 - 50,9	51,0 - 55,9	> 55,9
20-29	< 33,0	33,0 - 36,4	36,5 - 42,4	42,5 - 46,4	46,5 - 52,4	> 52,4
30-39	< 31,5	31,5 - 35,4	35,5 - 40,9	41,0 - 44,9	45,0 - 49,4	> 49,4
40-49	< 30,2	30,2 - 33,5	33,6 - 38,9	39,0 - 43,7	43,8 - 48,0	> 48,0
50-59	< 26,1	26,1 - 30,9	31,0 - 35,7	35,8 - 40,9	41,0 - 45,3	> 45,3
60+	< 20,5	20,5 - 26,0	26,1 - 32,2	32,3 - 36,4	36,5 - 44,2	> 44,2

Table Reference: The Physical Fitness Specialist Certification Manual, The Cooper Institute for Aerobics Research, Dallas TX, revised 1997 printed in Advance Fitness Assessment & Exercise Prescription, 3rd Edition, Vivian H. Heyward, 1998.p48

3.5. Response to exercise

There is considerable individual variability in the response to similar exercise training and genetic variants may play an important role in this variability. Some individuals are ‘low responders’ (improving their fitness only slightly following a specific exercise training) while others respond well or very well (‘high responders’).



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ADRB2	rs1042713 (G > A)	A	GA	Slight predisposition for exercise-induced fat loss
AGT	rs699 (A > G)	A	AA	Predisposition to improved response to exercise
IL1B	rs1143634 (G > A)	G	GG	No predisposition to inflammation and pain following acute exercise
LIPC	rs1800588 (C > T)	T	CC	No predisposition to an increase of HDL cholesterol in response to exercise
OPRM1	rs1799971 (A > G)	A	AA	Predisposition to normal sensitivity to perceived exertion and pain during exercise
PPARD	rs2267668 (G > A)	G	AA	Predisposition to fewer beneficial effects of exercise-centered lifestyle intervention
TNF-α	rs1800629 (G > A)	G	GG	Predisposition to improve performance in response to exercise

Physical characteristics

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4.1. Inflammation

Several genetic variations enhance the inflammatory response that allows slow repair of muscle damage after exercise.

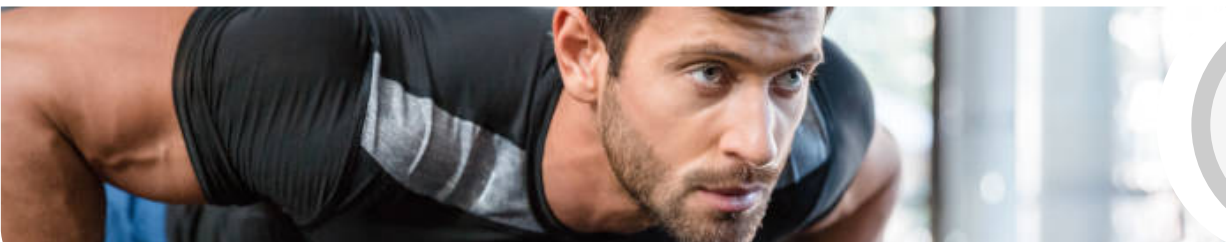


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GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
IL1B	rs1143634 (G > A)	G	GG	No predisposition to increased inflammation and pain perception following acute exercise
TNF-α	rs1800629 (G > A)	G	GG	Predisposition to normal levels of TNF-α and normal clearance of markers of muscle damage.

4.2. Oxidative stress

Oxidative stress occurs as a consequence of an imbalance between the formation of free radicals and the ability of the body to neutralize them on time. Regular exercise alleviates the negative effects caused by free radicals and offers many health benefits. However, physical performance is also known to induce oxidative stress, inflammation, and muscle fatigue. Polymorphisms in two important detoxification enzymes influence their activity and our exposure to oxidative stress during exercise.

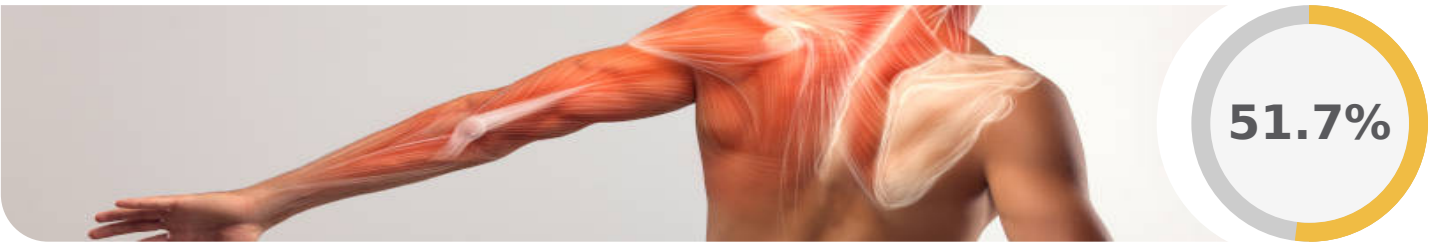


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GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
GSTP1	rs1695 (A > G)	G	AA	Predisposition to reduced GSTP1 activity leading to increased susceptibility to oxidative stress.
SOD2	rs4880 (A > G)	A	GG	Predisposition to reduced hydrogen peroxide detoxification and increased oxidative damage.

4.3. Muscle properties

Skeletal muscle contains a heterogeneous composition of different fiber types. This composition is an inherited trait that may partly predict athletic performance.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ACTN3	rs1815739 (C > T)	C	CT	Predisposition to increases in muscle strength and thickness in response to exercise
FTO	rs9939609 (T > A)	T	TT	Predisposition to normal proportion of slow-twitch muscle fibers
OPN	rs28357094 (T > G)	G	TT	Predisposition to normal muscle volume
PPARGC1A	rs8192678 (C > T)	C	TT	No predisposition to increase slow-twitch fiber content with exercise

4.4. Hypertension

The renin-angiotensin system plays a key role in maintaining blood pressure homeostasis as well as water and salt balance in the human body, significant factors associated with improved athletic performance, athlete fitness and exercise response. Being more active can lower blood pressure levels and reduce the risk of hypertension.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
AGT	rs699 (A > G)	A	AA	No additional risk of hypertension
EDN1	rs5370 (G > T)	G	GG	No additional risk of hypertension

4.5. Soft-tissue injuries

Increased physical activity puts a lot of pressure on tendons and muscles. This can cause injury to the tendons (tendinopathy) and muscles. Numerous scientific studies have linked a series of genes with an increased risk of tendinopathy and/or muscle injury.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
ACTN3	rs1815739 (C > T)	C	CT	No predisposition to exercise-induced muscle damage
COL5A1	rs12722 (C > T)	C	TT	Increased risk of ligament and tendon injuries
MMP-3	rs679620 (T > C)	T	TC	Reduced risk of Achilles tendon rupture
SOD2	rs4880 (A > G)	G	GG	Predisposition to normal levels of creatine kinase postexercise, an indirect marker of muscle damage.

4.6. Post-Exercise Recovery

On average, a muscle needs 24 to 48 hours to recover and repair the tissues that have been worn out during exercise or training. If the individual has slow clearance of lactate from the muscles, recovery time may be longer after intense physical exercise.



GENE	SNP (TRANSITION)	BENEFICIAL ALLELE	PATIENT GENOTYPE	GENOTYPE DESCRIPTION
AMPD1	rs17602729 (G > A)	G	GG	Predisposition to normal recovery time
SLC16A1	rs1049434 (A > T)	A	AT	Increased predisposition to muscle soreness and longer to recovery time

Understand genetics

5

Before reading this full report, please take a moment to read this overview to better understand the results and guide you on how best to use the results.



What is a genetic variant?

DNA is the "**instruction manual**" that governs the functioning of the human body. This "instruction manual" consists of long strands, containing many individual units called "**genes**", which in turn are made up of pairs of molecules called "**nucleotides**".

Each gene is comprised of thousands of combinations of four nucleotides that make up our genetic code: **A, T, C, and G**.

Nucleotides are the "spelling" that determine the function of each gene. The sequence formed by these bases varies between people. For example, at a specific location in a sequence, 80% of the population might have a "T", while the other 20% may have an "G". This specific difference of only one base, is called a **Single Nucleotide Polymorphism** or **SNP**.

SNPs are the most common type of genetic variation among people. Most SNPs have no effect on health or development. However, some create individual differences that influence how we look, and others create functional differences or affect biochemical reactions that are important for our long-term health and well-being.

What is an allele and a genotype?

An allele is each of two or more versions of a gene or SNP. The sum of the two alleles of an individual for a given position forms the genotype. If the two alleles are identical (example: AA), the genotype is homozygous for this position. If the two alleles are different (example: AG), the individual is heterozygous for this position. An allele that creates an individual difference (phenotype) can be dominant, semi-dominant, or recessive:

- **Dominant:** the effect of one allele on a heterozygous genotype completely masks the effect of the other.
- **Semi-dominant:** the phenotype of the heterozygous genotype is different from and often intermediate to the phenotypes of the homozygous genotypes.
- **Recessive:** the phenotype of the heterozygous genotype is not different from the phenotype of the homozygous wild type, two copies are necessary to obtain the phenotype studied.

What does it mean to have an unfavorable genotype?

Having an 'unfavourable' genotype does not mean that someone cannot play sports normally, it simply suggests that they may have more difficulties than someone with a more favorable genotype.

What is this genetic test based on?

This test is based on different genetic studies accepted by the scientific community. Section 8 indicates the main studies on which it is based.

Methodology

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How were the genetic variants selected and evaluated?

This test was developed by a multidisciplinary team of medical doctors, geneticists, and programmers, following highest quality standards. In particular, an expert team specialized in the curation of genetic variants reviewed each variant to ensure that selection, interpretation and impact of variants in the algorithms are based on the highest scientific evidence.

The following selection criteria were applied for classifying genetic variants:

Level 1A: Annotation for a variant in medical society-endorsed or implemented in a major health system.

Level 1B: Annotation for a variant where the preponderance of evidence shows an association. The association must be replicated in more than one cohort with significant p-values, and preferably will have a strong effect size.

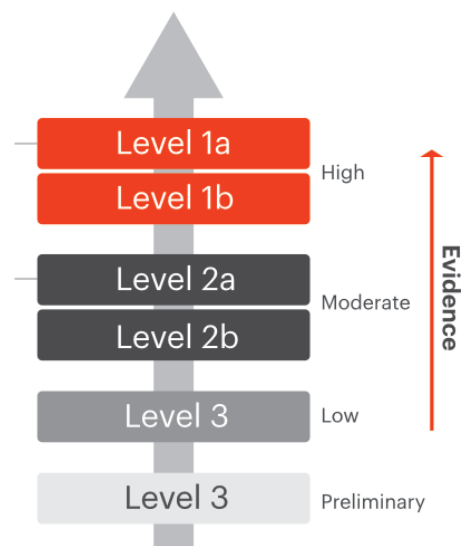
Level 2A: Annotation for a variant that qualifies for level 2B where the variant is within a Very Important known gene, so functional significance is more likely.

Level 2B: Annotation for a variant with moderate evidence of an association. The association must be replicated but there may be some studies that do not show statistical significance, and/or the effect size may be small.

Level 3: Annotation for a variant based on a single significant (not yet replicated) study or annotation for a variant evaluated in multiple studies but lacking clear evidence of an association.

Level 4: Annotation based on a case report, non-significant study or in vitro, molecular or functional assay evidence only.

Only genetic variants from level 1A to 2A were selected.



How was it analyzed?

The DNA was extracted from the buccal swab sample you provided and was analyzed by our clinical analysis laboratory. DNA was extracted using the KingFisher Flex® robotic extraction system (Thermo Fisher Scientific). The study of the genetic variants was performed by NGS (Next Generation Sequencing) using the Ion GeneStudio S5 system (Thermo Fisher Scientific).

What are the limits of this report?

- Each genetic marker tested is only one factor that predicts the probability of a particular result. However, your lifestyle, diet, and the environment you are exposed to all impact the ultimate effect predicted by your genes. These external factors cannot be considered in this report.
- For any questions about your health or personal clinical situation, you are advised to consult a qualified health professional.
- The information in this report is not used to diagnose genetic diseases or abnormalities, as it does not predict the risk and likelihood of certain genetic outcomes. It is also not intended to treat, diagnose, or cure any medical condition or disease.
- Our clinical laboratory has standard and effective procedures to protect against technical and operational problems. However, problems may occur in the shipment to the laboratory or in the handling of the sample, including, but not limited to, damage to the sample, mislabeling, and loss or delay in receiving the sample. In such cases, the medical laboratory may need to request a new sample.
- As with all medical laboratory tests, there is a small chance that the laboratory may provide inaccurate information.
- It is the responsibility of the professional who requests a test from us to guarantee the interested party appropriate genetic counseling in accordance with Law 14/2007, of July 3, on Biomedical Research.
- Fagron Genomics, S.L.U. declines all responsibility derived from the use and interpretation of the results of our tests by the requesting health professional.
- Fagron Genomics, S.L.U. does not access data identifying the patient from whom the sample comes, so it is also the responsibility of the requesting professional to comply with the applicable data protection regulations.

Analyzed genes

7

The detailed interpretation of the genetic study is attached. All evidence is supported by scientific articles indexed in PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>), identified in July 2023.

GENE (SNP)	DESCRIPTION
ACE (rs4343)	The angiotensin-converting enzyme (ACE) is a central component of the renin-angiotensin system, which controls blood pressure by regulating the volume of fluids in the body. ACE catalyses the conversion of angiotensin I to angiotensin II, which is a vasoconstrictive peptide, as well as the catabolism of bradykinin, which is a vasodilative peptide. The gene encoding ACE exhibits an insertion/deletion polymorphism (rs4343) characterized by either insertion (allele I) or deletion (allele D) of a 287 base-pair fragment. The absence (rs4343 G allele) of the D fragment Allele I is associated with higher circulating and tissue ACE levels [1] and consequently, with significantly higher angiotensin II and lower bradykinin levels. The ACE allele I, associated with a healthier cardiovascular system, is associated with increased VO2 peak [2], increased rates of muscle deoxygenation and reoxygenation [3], and improved endurance performance [4-5]. The absence (G) of the D fragment D allele of ACE increases the risk of developing cellulite [6].
ACTN3 (rs1815739)	α -actinin plays a decisive role in muscle contraction, acting as an anchor point or organizing center for the preformation of actin filaments. α -actinin 3 (ACTN3) is expressed only in fast-twitch muscle fibers, where it plays an important role in the generation of explosive and powerful muscle contractions. A common genetic variant in the ACTN3 gene (rs1815739 C>T) causes about 20% of the population with the TT genotype not to produce fast-twitch muscle fibres [7]. This mutation, also termed R577X, is not associated with any disease phenotype but affects athletic performance and muscle phenotypes. The C allele (R) is associated with increases in muscle strength and thickness in response to training [8-10], while the T allele (X) is associated with poorer sprint and power performance [11-15]. Carriers of the TT (XX) genotype are more susceptible to exercise-induced muscle damage [16-19].
ADRB2 (rs1042713)	The β 2 adrenergic receptor, also known as ADRB2, binds adrenaline, a hormone and neurotransmitter whose signaling mediates physiologic responses such as smooth muscle relaxation and bronchodilation. The Gly16Arg substitution (rs1042713 G>A) in ADRB2 has been shown to regulate the cardiopulmonary response to exercise [20]. rs1042713 influences elite endurance performance status, with the G (Gly) allele exerting an unfavourable effect [21-23]. Arg16 variant (A allele) showed a higher peak VO2max compared with Gly16 carriers [21, 24-25]. rs1042713 also influences exercise-induced fat loss and improvement of metabolic profile, with the G (Gly) allele exerting an unfavorable effect [26-28].

GENE (SNP)	DESCRIPTION
AGT (rs699)	The renin-angiotensin system is a hormonal system that regulates blood pressure, extracellular body volume, and sodium and potassium balance. The rs699 A>G polymorphism, also known as M235T, of the Angiotensinogen (AGT) gene is associated with higher plasma angiotensin concentrations [29-31], higher blood pressure and increased risk for hypertension [29, 32], which are higher in individuals homozygously carrying the G allele (G/G) or heterozygous (G/A) than in individuals carrying the A allele in homozygosity (AA). The G allele exerts a favourable effect on power performance by promoting skeletal muscle hypertrophy [12, 33-34]. It favourably modifies the influence of training or exercise on various health-related fitness parameters, such as cardiorespiratory endurance, blood pressure, and heart morphology [35-39].
AMPD1 (rs17602729)	The muscle-specific enzyme adenosine monophosphate deaminase 1 (AMPD1) is a very important regulator of muscle energy metabolism during exercise. The nonsense mutation (Gln12X, rs17602729) of the AMPD1 gene converts glutamine codon (CAA) into the premature stop codon (TAA), which results in the early interruption of protein synthesis. The AA genotype leads to a complete deficiency of the AMPD1 protein, which impairs AMP metabolism leading to muscle fatigue, weakness, and cramps, as well as a decreased exercise capacity, a lower response to exercise, and a lower relative increase in peak VO2 after aerobic training [40-41]. In contrast, the GG genotype has been associated with physical performance in power sports [42-46]. Carriers of the A allele have a limited training response of ventilatory phenotypes during maximal exercise [40, 47] and a reduced submaximal aerobic capacity [41].
AQP1 (rs1049305)	AQP1 encodes a small integral membrane protein which function as both a water channel and a non-selective monovalent cation channel when activated by intracellular cGMP. During osmotic stress, such as occurs during intense exercise, AQP1 facilitates water transfer from blood to muscle, provides osmotic protection, and promotes water reabsorption. The CG and CC genotypes of AQP1 rs1049305 polymorphism are associated with faster long-distance running performance [48- 52]. The presence of the G allele may result in a slower response to changes in osmotic gradients during exercise.
COL5A1 (rs12722)	The COL5A1 gene encodes the α1 chain of type V collagen that exerts a crucial role in the regulation of the size and configuration of other abundant fibrillar collagens supporting many tissues in the body, such as tendons, ligaments, and muscles [53]. A common C to T transition within the COL5A1 3' untranslated region (rs12722) may alter COL5A1 mRNA stability and thereby reduce the production of type V collagen [54]. Individuals with a TT genotype would have increased type V collagen production and thus favorably altered mechanical properties of tendons, which enhances endurance running ability [55-56]. The T allele (TT+TC) was associated with an increased risk of ligament and tendon injuries [57-59].

GENE (SNP)	DESCRIPTION
EDN1 (rs5370)	Endothelin 1 (EDN1) is a potent vasoconstrictor peptide and consequently, a key regulator of blood pressure. Carriers of the T allele of the EDN1 rs5370 polymorphism have an increased likelihood of having hypertension than GG genotypes, but this association is reduced by increasing physical activity or fitness level [60-61]. Carriers of the GT or TT genotypes are advised to engage in regular physical activity to minimize the risk of developing hypertension [62].
FTO (rs9939609)	The A allele of the common variation rs9939609 in the fat mass and obesity gene, FTO, was associated with increased fat mass, BMI, and risk of obesity. Homozygous AA carriers have a 3-fold higher risk for sarcopenia compared to T-allele carriers [63] and increased risk of muscle atrophy with aging [64]. The AA genotype is under-represented in athletes more reliant on a lean phenotype and associated with decreased proportion of slow-twitch muscle fibers. How ever it is over-represented in strength and heavier athletes [65].
GSTP1 (rs1695)	The glutathione S-transferase pi (GSTP1) gene plays an important regulatory role in detoxification, antioxidant damage and the onset of various diseases. GSTP1 rs1695 A>G (Ile105Val) is a widely studied GSTP1 polymorphism associated with modulation of GSTP1 enzyme activity. Carrying at least one copy of the GSTP1 G allele is associated with endurance [66-67] and significantly greater improvements in VO2max in response to aerobic training [68]. Children without a protective G allele showed an increased risk of new-onset asthma with increased participation in team sports [69].
HFE (rs1800562)	Human homeostatic iron regulator protein, also known as the HFE, participates in iron level sensing in the blood and in the modulation of hepcidin expression, a key regulator of the entry of iron into the circulation. Because intense activity may induce hepcidin synthesis, and thus decrease iron bioavailability, carriers of HFE mutations may have a competitive advantage in sports which require either a large capacity for oxygen transfer or high muscular performance. The HFE rs1800562 polymorphism accounts for ~85% of all cases of hemochromatosis. The hemochromatosis risk allele A has been associated with improved endurance performance and increased VO2 peak [70-72].

GENE (SNP)	DESCRIPTION
HIF1A (rs11549465)	The transcription factor HIF1A, induced in the presence of low oxygen concentrations, plays a key role in the physiological response to hypoxia. The T allele variant (Ser482) of the HIF1A rs11549465 polymorphism has shown increased transcriptional activity under hypoxic conditions compared to the wild-type C allele [73]. The T allele is associated with advantages in power sports, as well as a tendency to gain muscle mass and reduce body fat, which is consistent with the greater muscle mass of strength/power athletes [42, 74-76]. Genetic factors influencing the individual fibrogenic response to hypoxia in subcutaneous tissue may also play a crucial role in the pathogenesis of cellulite. The rare T allele of HIF1A rs11549465 has been shown to play a protective role against the development of cellulitis [8].
IL1B (rs1143634)	The Interleukin 1 beta (IL1B) is an important mediator of the inflammatory response. The A allele of the rs1143634 polymorphism in the IL1B gene is associated with the inflammation of skeletal muscle following acute exercise that may potentially affect exercise adaptations and pain perception [77-78].
LIPC (rs1800588)	Hepatic lipase (LIPC) hydrolyzes triglycerides and phospholipids present in circulating plasma lipoproteins and plays an important role in cholesterol homeostasis. The C allele of the LIPC rs1800588 (-514C>T) polymorphism is associated with reduced hepatic lipase activity and decreased serum HDL cholesterol levels [88]. Minor T allele carriers showed a significantly greater HDL-cholesterol increase in response to intensive sport-centered lifestyle intervention [88-91].
LRPPRC (rs10186876)	LRPPRC is a multifunctional protein considered essential for the assembly of the mitochondrial oxidative phosphorylation system, and therefore for energy production [92]. The rs10186876 LRPPRC polymorphism has been associated with handgrip strength, a widely used indicator of muscular fitness [93] and with weightlifting performance [94].

GENE (SNP)	DESCRIPTION
MMP-3 (rs679620)	The matrix metalloproteinase-3 (MMP-3) is involved in the breakdown of extracellular matrix proteins and during tissue remodeling in normal physiological processes. The CC genotype of rs679620 polymorphism in MMP3 is associated with an increased risk of Achilles tendinopathy, while the TT and TC genotypes were associated with a reduced risk to Achilles tendon rupture [14, 59, 79, 87].
OPN (rs28357094)	A polymorphism in the promoter of the osteopontin (OPN) gene (rs28357094) has been associated with multiple inflammatory states, severity of Duchenne muscular dystrophy and muscle size [96-97]. The G allele is associated with an increase in muscle volume [96, 98].
OPRM1 (rs1799971)	The rs1799971 polymorphism in the μ -opioid receptor gene (OPRM1) was related to changes in norepinephrine, lactate, and rate of perceived exertion and pain during exercise [99-100]. The general function of norepinephrine is to mobilize the brain and body for action. Athletes carrying at least one minor G allele were characterized by higher sensitivity when compared with AA wild-type homozygotes [78, 99-100].
PPARD (rs2267668)	The Peroxisome proliferator-activated receptor beta PPARD, also known as NR1C2, is a nuclear hormone receptor that governs a variety of biological processes. In muscle, PPARD expression is increased by exercise, resulting in increased oxidative (fat-burning) capacity and an increase in type I fibers. Genetic variations in PPARD determine mitochondrial function and change in aerobic physical fitness and insulin sensitivity during lifestyle intervention [101-102]. The PPARD rs2267668 G allele was associated with a negative response to aerobic training (decrease in VO2 peak) [102-103]. A allele carriers may enjoy fewer beneficial effects of exercise-centered lifestyle intervention on anthropometric indices and blood measurements [27, 104].

GENE (SNP)	DESCRIPTION
PPARG (rs1801282)	The peroxisome proliferator-activated receptors (PPARA, PPARG, PPARD) and their transcriptional coactivators (PPARGC1A, PPARGC1B) gene polymorphisms have been associated with muscle morphology, oxygen uptake, power output, and endurance performance. The alanine (Ala) variant of the PPARG gene Pro12Ala polymorphism (rs1801282 C>G) is associated with decreased receptor activity, improved insulin sensitivity [105], and increased body mass index in humans [106]. Carriers of the Ala allele show a better glycaemic response to exercise training [107], higher rates of skeletal muscle glucose uptake [108] and greater cross-sectional area of muscle fibres [109]. The G allele is associated with power athlete status [74, 110] with hypertrophic effect on muscle fibers [109]. Carriers of the CC (Pro/Pro) genotype are considered negative responders for aerobic training and have decreased glucose tolerance and insulin response) [110].
PPARGC1A (rs8192678)	PPARGC1A (or PGC-1α) regulates critical processes in muscle physiology, including mitochondrial biogenesis, lipid metabolism, angiogenesis, and the conversion of muscle fiber type towards slow-twitch type I fibers [103]. The PPARGC1A rs8192678 C>T polymorphism results in the substitution of glycine with serine in codon 482 (Gly482Ser). The minor T-allele (coding a serine residue) results in lower protein levels and reduced activity of the PPARGC1A protein [104]. The PPARGC1A rs8192678 polymorphism has been associated with athlete status in numerous publications [105]. Although the T (Ser) allele may impair aerobic capacity [95], it was shown to be more favorable for power athletes compared to controls [106-107]. The C (Gly) allele may be considered a beneficial factor for endurance performance [108-109]. T-allele carriers (Ser482) exhibit superior health benefits in increasing the VO2 max values when engaging in a physically active lifestyle compared to C-allele carriers (Gly482) [96, 110]. Carriers of minor T-allele (Ser) lacks a training-induced increase in the content of slow contracting oxidative fibers[111].
SLC16A1 (rs1049434)	During exercise, muscle fibers produce and accumulate lactate and hydrogen ions (H+) that may result in acute reduction in force and power (muscle fatigue). The H+-monocarboxylate cotransporter MCT1, encoded by the SLC16A1 gene, facilitates their removal by uptaking them into oxidative muscle fibers where lactate can be used as respiratory fuel [120-121]. A common MCT1 missense mutation (rs1049434) is associated with a strong reduction of lactate transport rate: Carriers of the T allele have a slower lactate transport rate and exhibit a higher lactate accumulation during high-intensity effort [122-124].
SOD2 (rs4880)	The superoxide dismutase 2 (SOD2) enzyme transforms toxic superoxide, a byproduct of the mitochondrial electron transport chain, into hydrogen peroxide and diatomic oxygen. SOD2 expression was found at least two-fold higher in the vastus lateralis muscle of elite athletes at baseline, indicating better antioxidant capacity [125-126]. The polymorphism rs4880 A>G (Val16Ala) of SOD2 has been shown to influence the antioxidative respons. The variant allele G reduces SOD2 catalytic activity [127-128]. The A allele was positively associated with athletic performance [12, 129], but also with increased creatine kinase (CK) postexercise [109, 130], an indirect marker of muscle damage.

GENE (SNP)	DESCRIPTION
TNF-α (rs1800629)	Tumor necrosis factor alpha (TNF-α) is a major regulator of inflammatory responses. The minor A allele of the rs1800629 polymorphism is associated with higher levels of TNF-α expression [131] and with impaired improvement of physical performance following physical activity [132-133]. Carriers of the GG genotype showed a greater decrease in C-reactive protein [134] and less elevated plasma creatine kinase following eccentric exercise [135] than the individuals with the A allele.
ZIC4 (rs11715829)	The rs11715829 polymorphism in the gene encoding the zinc finger protein ZIC4 is associated with gains in maximal O2 uptake (V̇o2max) after exercise training, with the major allele homozygotes (TT) gaining ~30% less than the minor allele carriers [136-137].

References

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1. Rigat et al., 1990 10.1172/JCI114844
2. Mägi et al., 2016 PMID: 27274666
3. Gasser et al., 2022 10.3389/fspor.2022.814975
4. Ma et al., 2013 10.1371/journal.pone.0054685
5. Ipekoglu et al., 2022 10.23736/S0022-4707.21.12417-X
6. Emanuele et al., 2010 10.1111/j.1468-3083.2009.03556.x
7. North et al., 1999 10.1038/7675
8. Gentil et al., 2011 PMID: 24149888
9. Clarkson et al., 2005 10.1152/japplphysiol.01139.2004
10. Norman et al., 2014 10.1152/japplphysiol.00557.2013
11. Alfred et al. 2011 10.1002/humu.21526
12. Weyerstraß et al. 2018 10.1016/j.jsams.2017.06.012
13. Tharabenjasin et al., 2019 10.1371/journal.pone.0217390
14. Appel et al., 2021 10.3389/fphys.2021.694411
15. Yang et al., 2003 10.1086/377590
16. Deuster et al., 2013 10.1007/s00421-013-2622-y
17. Vincent et al., 2009 10.1152/japplphysiol.01007.2009
18. Pimenta et al., 2012 10.1007/s00421-011-2109-7
19. Del Coso et al., 2017 10.1007/s00421-016-3507-7
20. Snyder et al., 2006 10.1113/jphysiol.2005.098558
21. Wolfarth et al., 2007 10.1016/j.metabol.2007.07.006
22. Tsianos et al., 2009 10.1152/japplphysiol.00780.2009
23. Sawczuk et al., 2013 10.1080/02640414.2013.786184
24. Wagoner et al., 2000 10.1161/01.res.86.8.834
25. Masschelein et al., 2015 10.1089/ham.2014.1083
26. Garenc et al., 2003 10.1038/oby.2003.88
27. Leońska-Duniec et al., 2018 10.1371/journal.pone.0202557
28. Camps et al., 2019 10.1016/j.gene.2019.100019
29. Jeunemaitre et al., 1992 10.1016/0092-8674(92)90275-h
30. Turgut et al., 2011 10.1007/s11033-010-0142-y
31. Chen et al., 2014 10.1177/1470320312465455
32. Kunz et al., 1999 10.1161/01.hyp.30.6.1331
33. Gómez-Gallego et al., 2014; 10.1007/s00421-009-1166-7
34. Zarębska et al., 2013 10.1519/JSC.0b013e31828155b5
35. Alves et al., 2009 10.1097/HJR.0b013e32832c5a8a
36. Karjalainen et al., 1999 10.1016/s0735-1097(99)00199-0
37. Rankinen et al., 2000 10.1152/ajpheart.2000.279.1.H368"
38. Rauramaa et al., 2002 10.1152/physiolgenomics.00050.2002.
39. Bruneau et al., 2015 10.1016/j.jsams.2015.05.009
40. Rico-Sanz et al., 2003 10.1152/physiolgenomics.00165.2002
41. Thomaes et al., 2011 10.1186/1471-2156-12-84
42. Cięszczyk et al., 2011 10.5604/945117
43. Cięszczyk et al., 2011 10.1055/s-0031-1283186
44. Cięszczyk et al., 2012 10.1080/02640414.2011.623710
45. Ginevičienė et al., 2014 10.1186/1471-2156-15-58
46. Murtagh et al., 2020 10.1371/journal.pone.0234458
47. Rubio et al. 2005 10.1152/japplphysiol.01371.2004
48. Martínez et al., 2009 10.2478/v10036-009-0039-9
49. Rivera et al., 2011 10.1055/s-0030-1268489
50. Rivera et al., 2019 10.1186/s40798-019-0213-0
51. Rivera et al., 2020 10.1186/s40798-020-00243-0
52. Saunders 2014 10.1080/02640414.2014.989535
53. Wenstrup et al., 2011 10.1074/jbc.M111.223693
54. Laguet et al., 2011 10.1016/j.matbio.2011.05.001
55. Posthumus et al., 2011 10.1249/MSS.0b013e3181f34f4d
56. Brown et al., 2011 10.1123/ijspp.6.4.485"
57. Alvarez-Romero et al., 2011 10.1080/17461391.2021.2011426
58. Lv et al., 2018 10.18632/oncotarget.23805
59. Guo et al., 2022 10.1186/s13018-022-03020-9
60. Asai et al., 2001 10.1161/hy1101.095333
61. Panoulas et al., 2008 10.1080/10623320802228708
62. Rankinen et al., 2007 10.1161/hypertensionaha.107.093609
63. Khanal et al., 2020a 10.1038/s41598-020-59722-9
64. Khanal et al., 2020b 10.3390/genes11121459
65. Guilherme et al., 2019 10.1519/JSC.0000000000003032
66. Gandhi et al., 2019 10.1016/j.heliyon.2019.e01928
67. Zabreska et al., 2016 10.1152/physiolgenomics.00014.2016
68. Zabreska et al., 2014 10.5604/20831862.1120932
69. Islam et al., 2009 10.1136/thx.2008.099366
70. Hermine et al., 2015 10.1016/j.biochi.2015.09.028
71. Thakkar et al., 2021 10.1249/MSS.0000000000002595
72. Thuc and Phong, 2021 <https://jjgastrohepto.org/wp-content/uploads/2021/09/JJGH-v7-1597.pdf>
73. Tanimoto et al., 2003 10.1093/carcin/bgg132
74. Drozdovska et al., 2013 10.5604/20831862.1059168
75. Gabbasov et al., 2013 10.1519/JSC.0b013e31827f06ae
76. Maciejewska-Karłowska et al., 2013 10.1371/journal.pone.0067172
77. Dennis et al., 2004 10.1113/jphysiol.2004.067876
78. Borsa et al., 2018 10.2147/JPR.S171498
79. Lulińska-Kuklik et al., 2019 PMC6370956
80. Ahmad et al., 2011 10.1161/CIRCGENETICS.110.957290
81. Brinkley et al., 2011 10.1152/japplphysiol.00567.2011
82. Huggins et al., 2013 10.1161/CIRCGENETICS.113.000042
83. Nassef et al., 2019 10.3390/genes10060440
84. Sasarman et al., 2015 10.1093/hmg/ddu468
85. Willems 2017 10.1038/ncomms16015
86. Kikuchi 2021 10.3390/genes13010025
87. Raleigh et al., 2009 10.1136/bjsm.2008.053892
88. Hoffman et al., 2013 10.1249/MSS.0b013e31828093c1
89. Pegoraro et al., 2011 10.1212/WNL.0b013e318207afef
90. Barfield et al., 2014 10.1093/hmg/ddu118
91. Karoly et al., 2012 10.1155/2012/540563
92. Ležnicka et al., 2018 10.1016/j.paid.2017.12.008
93. Thamer et al., 2008 10.1210/jc.2007-1209
94. Stephan et al., 2009 10.1210/jc.2006-1785
95. Petr et al., 2018 10.3390/ijms19051472
96. Nishida et al., 2015 10.2169/internalmedicine.54.3170
97. Deeb et al., 1998 10.1038/3099
98. Masus and Ye 2003 10.1136/jmg.40.10.773
99. Adamo et al., 2005 10.1007/s00125-005-1827-y
100. Vääntinen et al., 2005 10.1210/jc.2005-0101

101. Ahmetov et al., 2008 10.1007/s10517-009-0364-y
102. Petr et al., 2019 10.3390/ijms21010162
103. Lin et al., 2002 10.1038/nature00904
104. Prior et al., 2012 10.3233/DMA-2012-0894
105. Tharabenjasin et al., 2019 10.1371/journal.pone.0200967
106. Gineviciene et al., 2016 10.5604/20831862.1201051
107. Yang et al., 2021 10.3389/fgene.2021.726552"
108. Eynon et al., 2010 10.1111/j.1600-0838.2009.00930.x
109. Maciejewska et al., 2012 10.1080/02640414.2011.623709
110. Franks et al., 2003 10.1249/01.MSS.0000099109.73351.81
111. Steinbacher et al., 2015 10.1371/journal.pone.0123881
112. Pilegaard et al., 1999 10.1152/ajpendo.1999.276.5.E843
113. Dubouchaud et al., 2000 10.1152/ajpendo.2000.278.4.E571
114. Cupeiro et al., 2010 10.1016/j.jsams.2009.07.004
115. Cupeiro et al., 2012 10.1016/j.jsams.2012.03.009
116. Fedotovskaya et al., 2014 10.1123/ijsp.2013-0026
117. Brandauer et al., 2015 10.3389/fphys.2015.00085
118. Koltai et al., 2018 10.1016/j.redox.2018.07.022
119. Martin et al., 2009 10.1089/dna.2008.0788
120. Bastaki et al., 2006 10.1097/01.fpc.0000199498.08725.9c
121. Ben-Zaken et al. 2013 10.3109/10715762.2013.838627
122. Akimoto et al 2010 10.3109/10715760903494176
123. Karimi et al., 2009 10.1038/ejhg.2009.80
124. Li et al., 2016 10.1007/s11357-016-9909-y
125. Pereira et al., 2013 10.1007/s11357-013-9515-1
126. Kilpeläinen et al., 2010 10.1055/s-0030-1249686
127. Yamin et al. 2008 10.1007/s00421-008-0728-4
128. Sarzynski et al., 2017 10.1113/JP272559
129. Bouchard et al. 2011 10.1152/japplphysiol.00973.2010

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